

# ANMC Adult Rapid Blood Pathogen Identification Panel Guideline

(Last Updated: 5/2024)

ANMC utilizes an FDA approved test called the Blood Culture Identification 2 (BCID2) Panel performed on the FILMArray instrument. The test uses polymerase chain reaction (PCR) to amplify DNA targets from 3 groups of pathogens (gram positive bacteria, gram negative bacteria and yeast). It is performed directly following a positive blood culture and with an expansion in 2020 now allows rapid identification of 30 different pathogens (**Table 1**), including 10 genes responsible for vancomycin-resistant *Enterococci*, methicillin-resistant *Staphylococci*, CTX-M gene (ESBL), mcr-1 gene (colistin resistance), and five of the genes responsible for carbapenem-resistant *Enterobacterales*. In addition to detecting multiple species-specific assays, the panel also contains 3 genus specific assays (*Staphylococcus*, *Streptococcus* and *Enterobacterales*) that allows detection of pathogens for which there are no specific targets. A comprehensive list of all pathogens detected by BCID can be found in **Table 3**.

**Table 1: List of Pathogens/Resistance Genes Detected by Blood Pathogen Panel 2 (panel updated December 2021)**

Gram-Positive Bacteria	Gram-Negative Bacteria	Yeast	Resistance Markers
<i>Enterococcus faecalis</i> * <i>Enterococcus faecium</i> * <i>Listeria monocytogenes</i> <i>Staphylococcus</i> species <i>Staphylococcus aureus</i> <i>Staphylococcus epidermidis</i> * <i>Staphylococcus lugdunensis</i> * <i>Streptococcus</i> species <i>Streptococcus agalactiae</i> <i>Streptococcus pneumonia</i> <i>Streptococcus pyogenes</i>	<i>Acinetobacter calcoaceticus- baumannii</i> <i>Bacteroides fragilis</i> * <i>Enterobacterales</i> family <i>Enterobacter cloacae</i> complex <i>Escherichia coli</i> <i>Klebsiella aerogenes</i> * <i>Klebsiella oxytoca</i> <i>Klebsiella pneumoniae</i> group <i>Proteus</i> species <i>Salmonella</i> species* <i>Serratia marcescens</i> <i>Haemophilus influenzae</i> <i>Neisseria meningitidis</i> <i>Pseudomonas aeruginosa</i> <i>Stenotrophomonas maltophilia</i> *	<i>Candida albicans</i> <i>Candida auris</i> * <i>Candida glabrata</i> <i>Candida krusei</i> <i>Candida parapsilosis</i> <i>Candida tropicalis</i> <i>Cryptococcus neoformans/gattii</i> *	Carbapenemases <i>KPC</i> <i>IMP</i> * <i>OXA-48-like</i> * <i>NDM</i> * <i>VIM</i> * Colistin Resistance mcr-1* ESBL CTX-M* Methicillin (oxacillin) resistances <i>mecA/C</i> <i>mecA/C &amp; MREJ</i> * (MRSA) Vancomycin resistance <i>vanA/B</i>

\*Indicates new target on BCID2 panel

Immediately following a positive blood culture, the ANMC microbiology lab performs the BCID2. The results are typically available within 2 hours. It is important to note, that this test is NOT a standalone test. Blood samples with bacterial growth are still cultured and proper MIC testing is performed. The rapid reporting of pathogen identification allows for early escalation or de-escalation of antimicrobials to the most appropriate therapy while waiting for susceptibilities. A list of recommended antimicrobial treatment choices are outlined in **Table 3**.

The ANMC Antimicrobial Stewardship Program (ASP) developed these recommendations based upon the institutional antibiogram and national guidelines issued by the Infectious Diseases Society of America (IDSA). Relevant local susceptibility data is provided for gram-negative pathogens if activity for the antimicrobial is available. When both blood culture gram-stain and BCID2 results are available, current antimicrobial therapy should be evaluated in light of the clinical picture and adjusted to the most appropriate regimen. Additionally, when full susceptibility results become available therapy should again be evaluated and adjusted to the most appropriate narrow spectrum agent. It is important to note that certain infections are often polymicrobial in nature. In most cases, isolation of a single pathogen from the BCID2 should warrant narrowing of antimicrobial therapy, but should never result in over-narrowing. An example would be in complicated intra-abdominal infections where anaerobes are frequently present and therapy against these pathogens should generally be included until definitive cultures of the site of infection are available. Providers must use clinical judgment on a case-by-case basis.

**Table 2: Interpretations of *Staphylococcus* and *Enterobacterales* BCID2 Results**

Bacterial Marker	Result	Interpretation
<i>Staphylococcus</i> <i>S. aureus</i> , <i>S. epidermidis</i> , <i>S. lugdunensis</i>	Detected Not detected	Presumed Methicillin-resistant Coagulase-negative <i>Staphylococcus</i> species  The <i>mecA</i> analyte is not reported for non- <i>S. epidermidis</i> and <i>S. lugdunensis</i> coagulase-negative species (e.g. <i>S. hominis</i> , <i>S. simulans</i> , <i>S. capitis</i> , among others). Presume beta-lactam resistance.
<i>Staphylococcus</i> detected <i>S. epidermidis</i> <i>S. aureus</i> , <i>S. lugdunensis</i> <i>mecA</i>	Detected Detected Not Detected Not Detected	Methicillin-susceptible <i>Staphylococcus epidermidis</i>
<i>Staphylococcus</i> <i>S. aureus</i> <i>S. epidermidis</i> , <i>S. lugdunensis</i> MREJ and <i>mecA</i>	Detected Detected Not Detected Detected	Methicillin-resistant <i>S. aureus</i> (MRSA)
<i>Staphylococcus</i> <i>S. aureus</i> <i>S. epidermidis</i> , <i>S. lugdunensis</i> MREJ and <i>mecA</i>	Detected Detected Not Detected Not Detected	Methicillin-susceptible <i>S. aureus</i> (MSSA)
<i>Enterobacterales</i> <i>E. coli</i> CTX-M	Detected Detected Detected	Presumed ESBL producing <i>E. coli</i> *
<i>Enterobacterales</i> <i>Klebsiella pneumoniae</i> CTX-M	Detected Detected Not Detected	<i>Klebsiella pneumoniae</i> * (unlikely to have ESBL present)
<i>Enterobacterales</i> <i>Enterobacter cloacae</i> KPC	Detected Detected Detected	Presumed Carbapenem Resistant <i>Enterobacter cloacae</i> *
<i>Enterobacterales</i> All other species	Detected Not Detected	<i>Enterobacteriaceae</i> species lacking specific marker on the BCID2 panel (see Table 4)

\* There is very small chance that both the specific pathogen and another Enterobacteriaceae which cannot be not detected specifically by the BCID are present, but the therapy recommended should generally cover these pathogens as well

**Table 3: Blood Pathogen Panel Results and Recommended Empiric Therapy**

The table below contains interpretations of BCID2 data and recommended empiric antibiotic therapy for treating blood stream infections (BSIs) at ANMC. When the clinical picture and judgement dictates, patients who respond to narrow spectrum therapy do not always need to be escalated, even if this guideline recommends a broader spectrum agent. Similarly, patients who continue to deteriorate or are not clinically responding despite appropriate empirical antimicrobial therapy may require broader coverage. Allergies, organ dysfunction and history of multi-drug resistant organisms (MDROs) should always be considered prior to selecting empiric antimicrobial therapy.

## Gram Positive Organisms

Pathogen Detected		Preferred Empiric Therapy	Alternative Therapy	Comments/Considerations
<b><i>Enterococcus faecalis</i></b>		Ampicillin (regardless of VanA/B result)	Vancomycin (per pharmacy) If Van A/B positive- Linezolid or Daptomycin	Consider ID consult.
<b><i>Enterococcus faecium</i></b>	<i>Van A/B</i> Negative	Vancomycin (per pharmacy)	Linezolid	Consider ID consult.
	<i>Van A/B</i> Positive (VRE)	Linezolid	Daptomycin*	*High dose daptomycin (10mg/kg q24h) should be utilized for VRE blood stream infections
<b><i>Listeria monocytogenes</i></b>		Ampicillin	TMP/SMX in patients with severe beta-lactam allergy	Consider ID consult. GPR will not be run on BCID automatically. Please discuss with microbiology staff if concern for <i>Listeria</i> is present.
<b><i>Staphylococcus aureus</i></b>	<i>mecA/MREJ</i> Negative (MSSA)	Cefazolin	Nafcillin	Repeat Blood cultures (BCx) and formal ID consult recommended.
	<i>mecA/MREJ</i> Positive (MRSA)	Vancomycin (per pharmacy)	Daptomycin	Daptomycin should not be utilized when pneumonia is the primary source of infection.
<b><i>Staphylococcus sp. &amp; Staphylococcus epidermidis</i></b> ( <u>NOT</u> aureus or lugdunensis)	1 of 2 Positive	Probable contaminant. Consider withholding therapy. Do <u>not</u> need to routinely repeat BCx		<i>Staphylococcus epidermidis</i> <i>mecA</i> negative should be treated with cefazolin or nafcillin.
	2 of 2 Positive	Vancomycin (per pharmacy)	Daptomycin	
<b><i>Staphylococcus lugdunensis</i></b>	<i>mecA</i> Negative	Cefazolin	Nafcillin	Consider ID consult.
	<i>mecA</i> Positive	Vancomycin (per pharmacy)	Daptomycin	
<b><i>Streptococcus pyogenes</i></b> (Group A, Beta hemolytic Strep) and <b><i>Streptococcus agalactiae</i></b> (Group B, Beta hemolytic Strep)		Penicillin G <u>or</u> Ampicillin <u>or</u> Cefazolin	Vancomycin (per pharmacy)	Beta-hemolytic strep are routinely susceptible to penicillin
<b><i>Streptococcus pneumoniae</i></b>	Pneumonia	Ampicillin	Cefazolin Ceftriaxone	Continue vancomycin in patients with suspected CNS infection until susceptibilities are available.
	CNS Infection	Vancomycin (per pharmacy) + Ceftriaxone		
<b><i>Streptococcus</i> species (NOT Group A, NOT Group B, NOT <i>pneumoniae</i>)</b>	1 of 2 Positive	Possible contaminant, if no other source of infection, consider discussing with ID provider.		In severely ill or immunocompromised patients consider starting/continuing therapy until more definitive results are available.
	2 of 2 Positive	Ampicillin	Vancomycin (per pharmacy)	

Gram Negative Organisms (No resistance genes detected)			
Pathogen Detected	Preferred Empiric Therapy	Alternative Therapy	Comments/Considerations (Susceptibility data from 2023)
<i>Acinetobacter calcoaceticus-baumannii</i> complex	Meropenem 500mg IV q6h	Ampicillin/Sulbactam 3g IV q4h	Consider ID consult.
<i>Bacteroides fragilis</i>	Metronidazole	Piperacillin/Tazobactam	Usually part of underlying infection (i.e. intra-abdominal)
<b>Enterobacterales order ONLY</b>	Cefepime 2g IV q8h (extended infusion over 4 hours)	Levofloxacin Meropenem	See list of potential pathogens in <b>Table 4</b> Contact microbiology, ID, or Antimicrobial Stewardship with questions
<i>Enterobacter cloacae</i> *	Cefepime 2g IV q8h (extended infusion over 4 hours)	Levofloxacin Meropenem	<u>Cefepime</u> : 97% susceptible <u>Levofloxacin</u> : 93% susceptible <u>Meropenem</u> : 96% susceptible
<i>Escherichia coli</i>	Ceftriaxone	Piperacillin/Tazobactam	<u>Ceftriaxone</u> : 95% susceptible <u>Pip/Tazo</u> : 98% susceptible
<i>Klebsiella aerogenes</i> *	Cefepime 2g IV q8h (extended infusion over 4 hours)	Levofloxacin Meropenem	<u>Cefepime</u> : 100% susceptible <u>Levofloxacin</u> : 98% susceptible <u>Meropenem</u> : 100% susceptible
<i>Klebsiella oxytoca</i>	Ceftriaxone	Piperacillin/Tazobactam Levofloxacin	<u>Ceftriaxone</u> : 100% susceptible <u>Pip/Tazo</u> : 100% susceptible <u>Levofloxacin</u> : 100% susceptible
<i>Klebsiella pneumoniae</i> group	Ceftriaxone	Piperacillin/Tazobactam Levofloxacin	<u>Ceftriaxone</u> : 98% susceptible <u>Pip/Tazo</u> : 96% susceptible <u>Levofloxacin</u> : 92% susceptible
<i>Proteus</i> species	Ceftriaxone	Piperacillin/Tazobactam Levofloxacin	<u>Ceftriaxone</u> : 96% susceptible <u>Pip/Tazo</u> : 100% susceptible <u>Levofloxacin</u> : 94% susceptible
<i>Salmonella</i> species	Ceftriaxone		
<i>Serratia marcescens</i>	Ceftriaxone	Levofloxacin Meropenem	<u>Ceftriaxone</u> : 100% susceptible <u>Levofloxacin</u> : 100% susceptible <u>Meropenem</u> : 100% susceptible
<i>Haemophilus influenza</i>	Ampicillin/sulbactam <u>or</u> Ceftriaxone		De-escalate to ampicillin if beta-lactamase negative
<i>Neisseria meningitidis</i>	Ceftriaxone 2g IV q12h		<i>N. meningitidis</i> is associated with CNS infection. Rule out meningitis. Consider ID consult.
<i>Pseudomonas aeruginosa</i>	Cefepime 2g IV q8h (extended infusion over 4 hours)	Piperacillin/Tazobactam Ciprofloxacin	<u>Cefepime</u> : 99% susceptible <u>Pip/Tazo</u> : 96% susceptible <u>Ciprofloxacin</u> : 95% susceptible
<i>Stenotrophomonas maltophilia</i>	Sulfamethoxazole/Trimethoprim 15-20 mg/kg/day (TMP) in three divided doses	Levofloxacin	

\*AmpC chromosomal and inducible organisms may develop resistance during prolonged therapy with third-generation cephalosporins. Therefore, isolates that are initially susceptible may become resistant within three to four days after initiation of therapy.

Yeast Organisms		
Pathogen Detected	Preferred Empiric Therapy	Comments/Considerations
<i>Candida albicans</i>	Micafungin	Repeat Blood cultures (BCx) and formal ID consult recommended.
<i>Candida auris</i>	Micafungin	
<i>Candida glabrata</i>	Micafungin	
<i>Candida krusei</i>	Micafungin	
<i>Candida parapsilosis</i>	Micafungin	
<i>Candida tropicalis</i>	Micafungin	
<i>Cryptococcus neoformans/gattii</i>	Liposomal amphotericin B 3mg/kg daily	

Resistance Genes		
<i>IMP, KPC, OXA-48-like, NDM, VIM</i>	ID consult Ceftazidime/avibactam 2.5g IV q8h (infused over 3 hours) <b>PLUS</b> aztreonam 2g IV q8h (infused over 3 hours) <b>at the same time</b> as ceftazidime/avibactam	Markers for carbapenem-resistance in gram negative pathogens
<i>mcr-1</i>	Consider ID consult	Marker for colistin resistance
<i>CTX-M</i>	Meropenem 500 mg IV q6h preferred therapy	Marker for most common extended spectrum $\beta$ -lactamase (ESBL) found in gram-negative pathogens ESBLs hydrolyze expanded spectrum cephalosporins (ceftriaxone, cefepime) and piperacillin/tazobactam Negative results do not exclude the presence of other ESBL enzymes or other beta-lactamases
<i>mecA/C</i>	Vancomycin (pharmacy to dose)	<i>mecA/C</i> is a marker for methicillin/oxacillin resistance in non- <i>S. aureus</i> <i>Staphylococci</i> . Reported for <i>S. epidermidis</i> and <i>S. lugdunensis</i> .
<i>mecA/C</i> and <i>MREJ</i>	Vancomycin (pharmacy to dose)	<i>MREJ</i> is only evaluated in <i>S. aureus</i> and when present with <i>mecA/C</i> is specific for MRSA.
<i>vanA/B</i>	Linezolid 600 mg IV/PO q12h	Marker for vancomycin-resistant Enterococcus (VRE)

**Table 4: Pathogens Detected by Blood Culture Identification Panel 2**

Genus Specific Assay	Pathogens Detected by BCID2	Pathogens Not Detected By BCID2
<p><i>Enterococcus</i></p> <p>BCID included a genus level assay for <i>Enterococcus</i>. BCID2 does not include this genus assay, only including species specific detection for the 2 major species associated with blood stream infections.</p>	<ul style="list-style-type: none"> <li>• <i>E. faecium</i>*</li> <li>• <i>E. faecalis</i>*</li> </ul>	<ul style="list-style-type: none"> <li>• <i>E. avium</i></li> <li>• <i>E. casseliflavus</i></li> <li>• <i>E. durans</i></li> <li>• <i>E. gallinarum</i></li> <li>• <i>E. hirae</i></li> <li>• <i>E. dispar</i></li> <li>• <i>E. saccharolyticus</i></li> <li>• <i>E. raffinosus</i></li> <li>• <i>E. mundtii</i></li> </ul>
<p><i>Staphylococcus</i> genus</p>	<p>It is predicted that only 5 species will not be detected. Of those, only <i>S. equorum</i> has been reported in a clinical setting.</p>	<ul style="list-style-type: none"> <li>• <i>S. equorum</i></li> <li>• <i>S. fluerettii</i></li> <li>• <i>S. lentus</i></li> <li>• <i>S. muscae</i></li> <li>• <i>S. rostri</i></li> </ul>
<p><i>Streptococcus</i> genus</p> <p>Designed to detect most Viridans group species and non-Group A/B beta hemolytic streptococci.</p>	<p>All species within the <i>Streptococcus</i> genus should be amplified by one or more of the assays on the panel at positive blood culture levels. Some species may not be detected if present in a blood culture at low levels or if they have variant sequences (see right).</p>	<ul style="list-style-type: none"> <li>• <i>S. equi</i></li> <li>• <i>S. entericus</i></li> <li>• <i>S. halitosis</i></li> <li>• <i>S. hyovaginalis</i></li> <li>• <i>S. minor</i></li> <li>• <i>S. pantholopis</i></li> <li>• <i>S. oralis</i></li> <li>• <i>S. sobrinus</i></li> <li>• <i>S. suis</i></li> <li>• <i>S. uberis</i></li> </ul>
<p><i>Enterobacterales</i></p> <p>Designed to detect less common gram-negative bacteria within multiple families of the order Enterobacterales.</p> <p>Information about the detection of specific subspecies, strains, isolates, or serotypes of gram-negative bacteria is provided in the product instructions for use (Table 98 – Table 112) available at <a href="http://www.biofiredx.com/support/documents">www.biofiredx.com/support/documents</a>.</p>	<ul style="list-style-type: none"> <li>• <i>Cedeceae</i> spp.</li> <li>• <i>Citrobacter</i> spp.</li> <li>• <i>Cosenzaea</i> spp.</li> <li>• <i>Cronobacter</i> spp.</li> <li>• <i>Edwardsiella</i> spp (In silico predication)</li> <li>• <i>Enterobacter</i> spp.</li> <li>• <i>Escherichia</i> spp.</li> <li>• <i>Erwinia</i> spp.</li> <li>• <i>Hafnia</i> spp.</li> <li>• <i>Klebsiella</i> spp.</li> <li>• <i>Kluyvera</i> spp.</li> <li>• <i>Kosakonia</i> spp.</li> <li>• <i>Leclercia</i> spp.</li> <li>• <i>Lelliottia</i> spp.</li> <li>• <i>Mixta</i> spp.</li> <li>• <i>Morganella</i> spp.*</li> <li>• <i>Pantoea</i> spp.</li> <li>• <i>Phytobacter</i> spp.</li> <li>• <i>Plesiomonas</i> spp.</li> <li>• <i>Pluralibacter</i> spp.</li> <li>• <i>Providencia</i> spp.*</li> <li>• <i>Proteus</i> spp.</li> <li>• <i>Pseudoescherchia</i> spp</li> <li>• <i>Rahnella</i> spp.*</li> <li>• <i>Raoultella</i> spp.</li> <li>• <i>Salmonella</i> spp.*</li> <li>• <i>Serratia</i> spp.</li> <li>• <i>Sodalis</i> spp.</li> <li>• <i>Shigella</i> spp.</li> <li>• <i>Tatumella</i> spp.*</li> <li>• <i>Trabulsiella</i> spp.</li> <li>• <i>Yersinia</i> spp.*</li> <li>• <i>Yokanella</i> spp.</li> </ul>	<ul style="list-style-type: none"> <li>• <i>Providencia heimbachae</i></li> <li>• <i>Photorhabdus asymbiotica</i></li> <li>• <i>Arsenophonus nasoniae</i></li> </ul>

\*Indicates new species group detected by the BCID2 panel

#### References:

1. Barlam TF, Cosgrove SE, Abbo LM, et al. Implementing an Antibiotic Stewardship Program: Guidelines by the Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America. Clin Infect Dis. 2016;62(10):e51-e77. doi:10.1093/cid/ciw118
2. Timbrook TT, Morton JB, McConeghy KW, Caffrey AR, Mylonakis E, LaPlante KL. The Effect of Molecular Rapid Diagnostic Testing on Clinical Outcomes in Bloodstream Infections: A Systematic Review and Meta-analysis. Clin Infect Dis. 2017;64(1):15-23. doi:10.1093/cid/ciw649
3. Pardo J, Klinker KP, Borgert SJ, Butler BM, Giglio PG, Rand KH. Clinical and economic impact of antimicrobial stewardship interventions with the FilmArray blood culture identification panel. Diagn Microbiol Infect Dis. 2016;84(2):159-164. doi:10.1016/j.diagmicrobio.2015.10.023
4. Messacar K, Hurst AL, Child J, et al. Clinical Impact and Provider Acceptability of Real-Time Antimicrobial Stewardship Decision Support for Rapid Diagnostics in Children With Positive Blood Culture Results. J Pediatric Infect Dis Soc. 2017;6(3):267-274. doi:10.1093/jpids/piw047
5. BioFire® Blood Culture Identification 2 (BCID2) Panel [product labeling: instructions for use]. BioFire Diagnostics, Salt Lake City, UT; updated June 2020.